

Antitumor Sesquiterpenes from *Euonymus nanoides*

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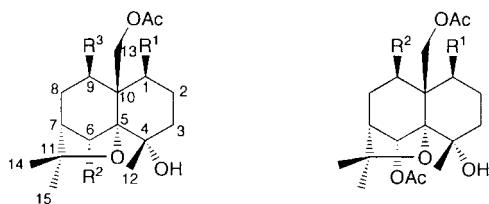
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The isolation, structure elucidation, and antitumor activity of four new sesquiterpene polyol esters, *i.e.*, of 6 α ,13-bis(acetyloxy)-9 β -(cinnamoyloxy)-1 β -(furan-3-ylcarbonyloxy)-4 α -hydroxy- β -dihydroagarofuran (**1**), 13-(acetyloxy)-9 β -(benzoyloxy)-4 α -hydroxy-1 β ,6 α -bis[(2-methylbutanoyloxy)- β -dihydroagarofuran (**2**), 1 β ,6 α ,13-tri(acetyloxy)-9 β -(cinnamoyloxy)-4 α -hydroxy- β -dihydroagarofuran (**3**), and 6 α ,13-bis(acetyloxy)-9 β -(benzoyloxy)-4 α -hydroxy-1 β -[(2-methylbutanoyloxy)- β -dihydroagarofuran (**4**), and of five known sesquiterpene polyol esters **5–9** from the seed oil of *Euonymus nanoides* LOES. are reported (β -dihydroagarofuran = octahydro-2,2,5a,9-tetramethyl-2H-3,9a-methano-1-benzoxepin).

Introduction. – Plants of the Celastraceae family comprise *ca.* 60 genera and 850 species worldwide [1]. Many of them have been used as traditional medicinal plants [2]. β -Dihydroagarofuran (= octahydro-2,2,5a,9-tetramethyl-2H-3,9a-methano-1-benzoxepin) sesquiterpene polyol esters and alkaloids are biologically active natural products found almost exclusively in species of the *Euonymus* (Celastraceae) genus [3–5]. The common structural features of this class of compounds are a terpene C₁₅ skeleton and a β -dihydroagarofuran moiety. Other structural elements can vary considerably. Pharmacological activities described for β -dihydroagarofuran sesquiterpenes include their cytotoxic [6], antitumor-promoting [7], immunosuppressive [8], insecticidal [9], and insect-antifeedant activities [10] and reversing multi-drug resistance in cancer cells [11]. Recently, several β -dihydroagarofuran sesquiterpenes were shown to have anti-HIV activity [12]. As part of our interest in β -dihydroagarofuran sesquiterpene polyol esters with antitumor activity, we investigated the chemical constituents of the seed oil of *Euonymus nanoides* LOES., which showed antiproliferative activity in the A-549, HL-60, BEL-7402, and P-388 cell-line antitumor screens. We now wish to report the isolation, structure determination, and biological activity of the four new compounds **1–4** and of the five known compounds **5–9** from the seed oil of *E. nanoides*.

Results and Discussion. – Column chromatography (CC), vacuum liquid chromatography (VLC), thin layer chromatography (TLC), and reversed-phase column chromatography (*RP-18*) of the acetone extract of the seed of *E. nanoides* yielded compounds **1–9**. The structures of compounds **5–9** were determined by spectral data as 1 β ,2 β ,6 α -tris(acetyloxy)-9 α -(benzoyloxy)-4 α -hydroxy-13-[(2-methylbutanoyloxy)- β -dihydroagarofuran¹) (**5**) [13], 2 β ,6 α -bis(acetyloxy)-1 β -[(furan-3-ylcarbonyloxy)-4 α -

¹) Arbitrary numbering (see *Formula 1*); for systematic names of **1–4**, see *Exper. Part*

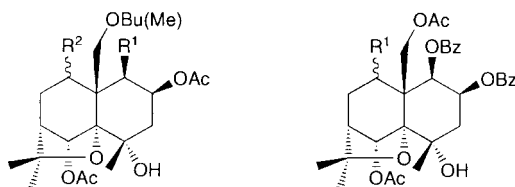


1) R¹ = FuO, R² = AcO, R³ = CinO

2) R¹ = R² = Bu(Me)O, R³ = BzO

3) R¹ = AcO, R² = CinO

4) R¹ = Bu(Me)O, R² = BzO



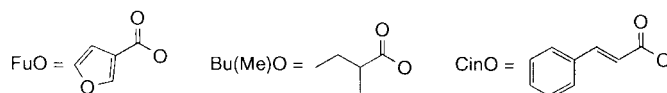
5) R¹ = AcO, R² = α -BzO

6) R¹ = FuO, R² = β -Bu(Me)O

9) R¹ = BzO, R² = α -Bu(Me)O

7) R¹ = α -AcO

8) R¹ = β -BzO



hydroxy-9 β ,13-bis-[(2-methylbutanoyl)oxy]- β -dihydroagarofuran¹) (**6**) [14], 6 α ,9 α ,13-tris(acetyloxy)-1 β ,2 β -bis(benzoyloxy)-4 α -hydroxy- β -dihydroagarofuran¹) (**7**) [14], 6 α ,13-bis(acetyloxy)-1 β ,2 β ,9 β -tris(benzoyloxy)-4 α -hydroxy- β -dihydroagarofuran¹) (**8**) [15], 2 β ,6 α -bis(acetyloxy)-1 β -(benzoyloxy)-4 α -hydroxy-9 α ,13-bis[(2-methylbutanoyl)oxy]- β -dihydroagarofuran¹) (**9**) [14]; similarly, the structures of the new compounds **1–4** were established.

Compound **1** was isolated as an optically active yellow oil. The HR-MS of **1** exhibited a quasi-molecular-ion peak at m/z 611.2494 ($[M + H]^+$), corresponding to the molecular formula C₃₃H₃₈O₁₁. The IR spectrum revealed a characteristic ester absorption at 1732 cm⁻¹ and a free OH absorption at 3432 cm⁻¹. The complete assignments of the ¹H- and ¹³C-NMR data of **1** were successfully carried out by ¹H,¹H COSY, HMQC, and HMBC experiments (Tables 1 and 2). Thus, compound **1** was identified as 6 α ,13-bis(acetyloxy)-9 β -(cinnamoyloxy)-1 β -[(furan-3-ylcarbonyl)oxy]-4 α -hydroxy- β -dihydroagarofuran¹).

The NMR spectra of **1** suggested the presence of two acetate moieties (δ (H) 1.79, 2.20 (2s, 2 Me); δ (C) 20.6 (*q*), 21.3 (*q*), 169.7 (*s*), 170.8 (*s*)), a cinnamate function (δ (H) 6.38 (*d*, $J = 15.6$ Hz, PhCH=CH), 7.40 (*m*, 2 H, Ph), 7.47 (*m*, 1 H, Ph), 7.55 (*m*, 2 H, Ph), 7.70 (*d*, $J = 15.6$ Hz, PhCH=CH); δ (C) 117.7 (*d*), 128.8 (*2d*), 129.7 (*2d*), 130.5 (*d*), 134.2 (*s*), 145.8 (*d*), 165.8 (*s*)), a furan-3-carboxylate moiety (δ (H) 6.87 (*d*, $J = 1.2$ Hz, 1 H, CH=CH), 7.41 (*d*, $J = 1.2$ Hz, 1 H, CH=CH), 8.27 (*s*, CH=C); δ (C) 109.7 (*d*), 118.9 (*s*), 144.0 (*d*), 148.4 (*d*), 161.9 (*s*)), and a free OH group (δ (H) 2.71 (*s*)). The ¹H-NMR spectrum (400 MHz, CDCl₃) of **1** showed the presence of three tertiary Me groups at δ 1.34 (*s*, Me(12)), 1.46 (*s*, Me(14)), and 1.49 (*s*, Me(15))¹). The ¹H,¹H

Table 1. $^1\text{H-NMR}$ Data (400 MHz) of Compounds **1–4** in CDCl_3 , $\delta(\text{H})$ in ppm, J in Hz.

	1	2	3	4
H–C(1)	5.66 (<i>d</i> , $J=4.0$)	5.57 (<i>d</i> , $J=4.0$)	5.48 (<i>d</i> , $J=3.2$)	5.59 (<i>d</i> , $J=3.6$)
CH ₂ (2)	2.45, 2.07 (<i>2m</i>)	2.48, 2.08 (<i>2m</i>)	2.40, 2.01 (<i>2m</i>)	2.46, 2.11 (<i>2m</i>)
CH ₂ (3)	1.15, 0.90 (<i>2m</i>)	1.50, 2.01 (<i>2m</i>)	1.20, 0.86 (<i>2m</i>)	1.50, 2.02 (<i>2m</i>)
H–C(6)	5.67 (<i>s</i>)	5.69 (<i>s</i>)	5.58 (<i>s</i>)	6.15 (<i>s</i>)
H–C(7)	2.31 (<i>m</i>)	2.33 (<i>m</i>)	2.21 (<i>m</i>)	2.30 (<i>m</i>)
CH ₂ (8)	2.31, 2.11 (<i>2m</i>)	2.31, 2.15 (<i>2m</i>)	2.25, 2.11 (<i>2m</i>)	2.32, 2.17 (<i>2m</i>)
H–C(9)	5.28 (<i>d</i> , $J=6.8$)	5.34 (<i>d</i> , $J=6.8$)	5.21 (<i>d</i> , $J=6.8$)	5.34 (<i>d</i> , $J=6.4$)
Me(12)	1.34 (<i>s</i>)	1.34 (<i>s</i>)	1.31 (<i>s</i>)	1.33 (<i>s</i>)
CH ₂ (13)	5.04, 4.42 (<i>2d</i> , each $J=12.8$)	4.87, 4.52 (<i>2d</i> , each $J=12.8$)	4.75, 4.44 (<i>2d</i> , each $J=12.8$)	4.79, 4.56 (<i>2d</i> , each $J=12.8$)
Me(14)	1.46 (<i>s</i>)	1.43 (<i>s</i>)	1.43 (<i>s</i>)	1.42 (<i>s</i>)
Me(15)	1.49 (<i>s</i>)	1.46 (<i>s</i>)	1.46 (<i>s</i>)	1.44 (<i>s</i>)

Table 2. $^{13}\text{C-NMR}$ Data (100 MHz) of Compounds **1–4** in CDCl_3 , $\delta(\text{C})$ in ppm.

	1^a	2^a	3^a	4^a
CH(1)	70.1	70.0	70.1	70.0
CH ₂ (2)	31.0	31.1	30.9	31.0
CH ₂ (3)	40.5	41.7	40.5	40.5
C(4)	69.5	69.4	69.5	69.4
C(5)	89.7	89.6	89.7	89.8
CH(6)	69.1	70.2	69.0	68.5
CH(7)	43.3	43.4	43.2	43.3
CH ₂ (8)	33.6	33.5	33.4	33.3
CH(9)	69.0	68.3	68.6	68.1
C(10)	50.8	51.2	50.9	51.3
C(11)	83.7	83.8	83.6	83.8
Me(12)	30.0	29.9	29.9	29.9
CH ₂ (13)	66.3	66.2	65.6	65.6
Me(14)	25.3	26.5	25.0	25.1
Me(15)	24.2	24.4	24.2	24.3

^a) Attached protons by DEPT.

COSY plot exhibited signals at δ 5.66 (*d*, $J=4.0$ Hz, H–C(1)), 5.67 (*s*, H–C(6)), and 5.28 (*d*, $J=6.8$ Hz, H–C(9)) assigned to CH groups bearing an ester group, while signals at δ 4.42 (*d*, $J=12.8$ Hz, H_a–C(13)) and δ 5.04 (*d*, $J=12.8$ Hz, H_b–C(13)) were assigned to the a CH₂ group bearing an ester group. The $^{13}\text{C-NMR}$ spectrum (100 MHz, CDCl_3) of **1** showed 3 Me (δ 24.2, 25.3, and 30.0), 3 CH₂ (δ 31.0, 33.6, and 40.5), 1 OCH₂ (δ 66.3), 1 CH (δ 43.3), 3 OCH (δ 69.0, 69.1, and 70.1), 1 quaternary C-atom (δ 50.8), and 3 quaternary C-atoms attached to an O-atom (δ 69.5, 83.7, and 89.7), whose chemical shifts were very similar to those of reported β -dihydroagarofuran sesquiterpene polyol esters [16][17]. The above data suggested that **1** was a β -dihydroagarofuran sesquiterpene substituted with two acetyloxy, one (furan-3-ylcarbonyl)oxy, one cinnamoyloxy, and one free OH groups. The $^1\text{H},^{13}\text{C}$ long-range correlation spectrum (HMBC) confirmed the location of the ester groups. Thus, the C=O signals at $\delta(\text{C})$ 161.9 and 165.8 were correlated with the signals at $\delta(\text{H})$ 5.66 (H–C(1)) and 5.28 (H–C(9)), respectively, revealing that the (furan-3-ylcarbonyl)oxy group was located at C(1) and the cinnamoyloxy group at C(9). Similarly, the acetyloxy groups were positioned at C(6) and C(13) because the C=O signals at $\delta(\text{C})$ 169.7 and 170.6 were correlated with the proton signals of H–C(6) and CH₂(13), respectively. As usually found in this class of compounds, H–C(1) and H–C(6) were axially positioned [9][18]. From the NOESY spectrum of **1**, the strong correlation between H–C(1) and H–C(9) indicated also an axial conformation for H–C(9)).

Compound **2** was isolated as an optically active yellow oil that analyzed for $C_{34}H_{48}O_{10}$ by HR-MS (m/z 639.3134 ($[M+Na]^+$)). The IR spectrum revealed a characteristic ester absorption at 1727 cm^{-1} and a free-OH absorption at 3432 cm^{-1} . In accord with the ^1H - and ^{13}C -NMR data (Tables 1 and 2), compound **2** was elucidated as 13-(acetyloxy)-9 β -(benzoyloxy)-4 α -hydroxy-1 β ,6 α -bis[(2-methylbutanoyl)oxy]- β -dihydroagarofuran¹).

The NMR spectra of **2** suggested the presence of two 2-methylbutanoate moieties ($\delta(\text{H})$ 0.55 (*t*, 2 *MeCH*₂), 0.80 (*d*, $J = 6.8\text{ Hz}$, 2 *MeCH*), 0.92 (*m*, *MeCH*₂), 1.20 (*m*, *MeCH*₂), 1.90 (*m*, *MeCH*₂*CH*), 2.00 (*m*, *MeCH*₂*CH*); $\delta(\text{C})$ 11.2 (*q*), 11.6 (*q*), 15.4 (*q*), 16.5 (*q*), 25.1 (*t*), 25.4 (*t*), 40.6 (*d*), 40.8 (*d*), 174.4 (*s*), 175.2 (*s*)), an acetate moiety ($\delta(\text{H})$ 2.21 (*s*, *Me*); $\delta(\text{C})$ 21.4 (*q*), 170.5 (*s*)), a benzoate function ($\delta(\text{H})$ 7.45 (*t*, 2 *H*, *Ph*), 7.57 (*t*, 1 *H*, *Ph*), 8.05 (*d*, $J = 7.6\text{ Hz}$, 2 *H*, *Ph*); $\delta(\text{C})$ 128.3 (*2d*), 129.4 (*s*), 130.2 (*2d*), 133.3 (*d*), 165.4 (*s*)), and a free OH group ($\delta(\text{H})$ 2.68 (*s*)). The NMR data for the parent ring system was very similar to those of **1**, suggesting that **2** also contains the 1,4,6,9,13-pentasubstituted β -dihydroagarofuran skeleton, and the locations of the protons were confirmed by the ^1H , ^1H COSY plot. As for **1**, the free OH group at C(4) was in equatorial position, and the locations of the ester groups were apparent from the HMBC spectrum, which showed cross-peaks between H–C(9) ($\delta(\text{H})$ 5.34, *d*, $J = 6.8\text{ Hz}$) and the C=O group at $\delta(\text{C})$ 165.4 of the benzoate moiety between CH₂(13) ($\delta(\text{H})$ 4.52, 4.87, *2d*, each $J = 12.8\text{ Hz}$, 2 *H*) and the C=O group at $\delta(\text{C})$ 170.5 of the acetate function, and between H–C(1) ($\delta(\text{H})$ 5.57, *d*, $J = 4.0\text{ Hz}$) and H–C(6) ($\delta(\text{H})$ 5.69, *s*) and the C=O groups at $\delta(\text{C})$ 175.2 and 174.4, respectively, of the two 2-methylbutanoate moieties. In the NOESY of **2**, a strong correlation between H–C(1) and H–C(9) was observed, which further supported that H–C(9) was in axial position.

Compounds **3** and **4** were obtained as yellow oils. The conclusions drawn from the HR-MS, ^1H -NMR, NOESY difference measurement, and ^{13}C -NMR of **3** and **4** were confirmed by ^1H , ^1H COSY, HMQC, and HMBC experiments (Table 1 and 2) and by comparison with data of **1** and **2**. Therefore, compound **3** was elucidated as 1 β ,6 α ,13-tris(acetyloxy)-9 β -(cinnamoyloxy)-4 α -hydroxy- β -dihydroagarofuran¹) and compound **4** as 6 α ,13-bis(acetyloxy)-9 β -(benzoyloxy)-4 α -hydroxy-1 β -[(2-methylbutanoyl)oxy]- β -dihydroagarofuran¹).

Compound **3** analyzed for $C_{30}H_{38}O_{10}$ by HR-MS (m/z 559.2534 ($[M+H]^+$)). The NMR spectra suggested the presence of three acetate functions ($\delta(\text{H})$ 1.79, 2.12, 2.16 (3*s*, 3 *Me*); $\delta(\text{C})$ 20.6 (*q*), 21.1 (*q*), 21.3 (*q*), 169.7 (*s*), 170.2 (*s*), 170.4 (*s*)), a cinnamate moiety ($\delta(\text{H})$ 6.35 (*d*, $J = 16.0\text{ Hz}$, *PhCH=CH*), 7.38 (*m*, 2 *H*, *Ph*), 7.52 (*m*, 1 *H*, *Ph*), 7.55 (*m*, 2 *H*, *Ph*), 7.67 (*d*, $J = 16.0\text{ Hz}$, *PhCH=CH*); $\delta(\text{C})$ 117.7 (*d*), 128.2 (*2d*), 128.8 (*2d*), 130.6 (*d*), 134.2 (*s*), 145.7 (*d*), 165.8 (*s*)), and a free OH group ($\delta(\text{H})$ 2.66 (*s*)). The NMR data for the parent ring system were very similar to those of **1** and **2**, suggesting that **3** also has the 1,4,6,9,13-pentasubstituted- β -dihydroagarofuran skeleton, and the locations of the protons were confirmed by the ^1H , ^1H COSY plot. As for **1**, the positions of the ester groups were determined from the HMBC spectrum, which showed cross-peaks between H–C(9) ($\delta(\text{H})$ 5.21, *d*, $J = 6.8\text{ Hz}$) and the C=O group at $\delta(\text{C})$ 165.8 of the cinnamate moiety, between CH₂(13) ($\delta(\text{H})$ 4.44, 4.75 *2d*, each $J = 12.8\text{ Hz}$, 2 *H*), H–C(1) ($\delta(\text{H})$ 5.48, *d*, $J = 3.2\text{ Hz}$) and H–C(6) ($\delta(\text{H})$ 5.58, *s*) and the C=O groups at $\delta(\text{C})$ 170.4, 169.7, and 170.2, respectively, of the acetate functions. In the NOESY of **3**, the correlation between H–C(1) and H–C(9) indicated the axial position of H–C(9).

Compound **4** analyzed for $C_{31}H_{42}O_{10}$ by HR-MS (m/z 575.2860 ($[M+H]^+$)). The NMR spectra suggested the presence of a 2-methylbutanoate group ($\delta(\text{H})$ 0.54 (*t*, *MeCH*₂), 0.79 (*d*, $J = 6.8$, *Me(CH)*), 0.91 (*m*, 1 *H*, *MeCH*₂), 1.18 (*m*, 1 *H*, *MeCH*₂), 2.00 (*m*, *MeCH*₂*CH*); $\delta(\text{C})$ 11.1 (*q*), 15.7 (*q*), 25.3 (*t*), 40.6 (*d*), 174.3 (*s*)), two acetate moieties ($\delta(\text{H})$ 2.08, 2.19 (2*s*, 2 *Me*); $\delta(\text{C})$ 21.0 (*q*), 21.2 (*q*), 169.4 (*s*), 170.4 (*s*)), a benzoate group ($\delta(\text{H})$ 7.44 (*t*, 2 *H*, *Ph*), 7.56 (*t*, 1 *H*, *Ph*), 8.03 (*d*, $J = 7.6\text{ Hz}$, 2 *H*, *Ph*); $\delta(\text{C})$ 128.2 (*2d*), 129.3 (*s*), 130.1 (*2d*), 133.5 (*d*), 165.3 (*s*)), and a free OH group ($\delta(\text{H})$ 2.68 (*s*)). The assignments of the other ^1H - and ^{13}C -NMR signals of **4** were successfully carried out with ^1H , ^1H COSY and HMQC experiments by comparison with data of **1–3**, and the positions of the ester groups were determined from the HMBC spectrum, which showed cross-peaks between H–C(9) ($\delta(\text{H})$ 5.34, *d*, $J = 6.4\text{ Hz}$) and the C=O group at $\delta(\text{C})$ 165.3 of the benzoate moiety, between H–C(1) ($\delta(\text{H})$ 5.69, *d*, $J = 3.6\text{ Hz}$), and the C=O group at $\delta(\text{C})$ 174.3 of the 2-methylbutanoate function, and between CH₂(13) (δ 4.56, 4.79, *2d*, each $J = 12.8\text{ Hz}$, 2 *H*), and H–C(6) ($\delta(\text{H})$ 6.15, *s*) and the C=O group at

$\delta(\text{C})$ 170.4 and 169.4, respectively, of the two acetate functions. The NOESY of **4** indicated the axial position of H–C(9).

All new and known compounds were tested for *in vitro* antitumor activity against three human-tumor cell lines, *i.e.*, A-549 (lung carcinoma), HL-60 (leukemia neoplasm), BEL-7402 (liver carcinoma), and one mouse-tumor cell line, *i.e.*, P-388 (leukemia neoplasm). IC_{50} Values were determined for compounds **1–9** (Table 3). These results show that the compounds are able to inhibit tumoral activity, with IC_{50} values below 100 μM .

Table 3. *in vitro* Antitumor Activities (IC_{50} [μM]) of Compounds **1–9** and VP-16.

	P-388	HL-60	A-549	BEL-7402
1	64.41	39.72	83.05	46.45
2	72.24	29.30	> 100	52.72
3	> 100	91.12	> 100	–
4	68.06	45.59	> 100	62.02
5	> 100	98.24	–	> 100
6	13.26	8.68	> 100	12.08
7	91.53	54.46	> 100	74.13
8	80.86	17.48	–	41.13
9	71.83	12.05	> 100	6.23
VP-16	5.89	5.08	8.89	13.17

Experimental Part

General. Column chromatography (CC): silica gel (200–300 mesh; *Quingdao of China*); reversed-phase, *RP-18* short column, 25×250 mm, *Merck*. Vacuum liquid chromatography (VLC): silica gel 60 (35–70 mesh; *Merck*). TLC: precoated silica gel 60, *F 254* plates (*Merck*); detection by UV light or by heating after spraying with 5% H_2SO_4 in EtOH. Optical rotation: *Perkin-Elmer-341* polarimeter. IR: *Nicolet-AVATAR-360-FT-IR* spectrometer; on KBr disks; in cm^{-1} . UV: *Shimadzu-UV-260* spectrometer; in nm. ^1H - and ^{13}C -NMR: at 400 and 100 MHz, resp.; *Bruker-Avance-400* spectrometer; chemical shifts δ in ppm, coupling constants J in Hz; SiMe_4 as internal standard; 2D NMR experiments included ^1H , ^1H COSY, HMQC, NOESY, DEPT, and HMBC. MS: *HP-5988-MS* and *APEXTMII-Bruker-4.7TAS* spectrometer.

Plant Material. The seed of *Euonymus nanoides* LOES. was collected in Luqu country, Gansu province of China in October 1997, and identified by Prof. J. Zh. Sun of the Department of Biology, Lanzhou University. A voucher specimen (No. 971001) is deposited at the Department of Biology, Lanzhou University.

Extraction and Isolation. Dried, powdered seeds (1.2 kg) of *E. nanoides* were extracted $3 \times$ for 7 d with acetone by percolation at r.t. to give a residue (102.8 g) after evaporation. This residue was separated by VLC and CC with a petroleum ether (60–90°)/acetone gradient. Compounds **1**, **3**, and **4** were eluted with petroleum ether/acetone 3 : 1, and **2** with petroleum ether/acetone 5 : 1. TLC purification of the crude compounds gave **1** (petroleum ether/acetone 3 : 2; 15.0 mg), **2** (petroleum ether/acetone 5 : 2; 7.0 mg), **3** (petroleum ether/acetone 2 : 1; 59.1 mg), and **4** (petroleum ether/acetone; 2 : 1 33.2 mg). Compounds **5–9** were eluted with petroleum ether/acetone 1 : 1 and purified by CC (*RP-18* MeOH/ H_2O 6 : 1 \rightarrow 2 : 1): **5** (5.0 mg), **6** (5.0 mg), **7** (6.7 mg), **8** (4.8 mg), and **9** (6.3 mg).

6 α ,13-Bis(acetyloxy)-9 β -(cinnamoyloxy)-1 β -[(furan-3-ylcarbonyloxy)-4 α -hydroxy- β -dihydroagarofuran (= Furan-3-carboxylic Acid (3R,5R,5aS,6S,9S,9aS,10R)-10-(Acetyloxy)-5a-[(acetyloxy)methyl]-octahydro-9-hydroxy-2,2,9-trimethyl-2H-3,9a-methano-1-benzoxepin-6-yl Ester; **1):** Yellow oil. $[\alpha]_D^{20} = +8.5$ ($c = 0.75$, CHCl_3). UV (MeOH): 205, 217, 251, 280. IR (KBr): 3432, 2931, 1732, 1638, 1230, 1159, 1023, 877, 721. ^1H - and ^{13}C -NMR (CDCl_3): Table 1 and 2. EI-MS: 550 (0.7, $[M - \text{AcOH}]^+$), 462 (15.0, $[M - \text{CinO}]^+$), 380 (64.0, $[M + \text{H} - 2\text{AcO} - \text{FuO}]^+$), 230 (100). HR-MS: 611.2494 ($[M + \text{H}]^+$, $\text{C}_{33}\text{H}_{39}\text{O}_{11}$; calc. 611.2487).

13-(Acetyloxy)-9 β -(benzoyloxy)-4 α -hydroxy-1 β ,6 α -bis[(2-methylbutanoyl)oxy]- β -dihydroagarofuran (=2-Methylbutanoic Acid (3R,5R,5aS,6S,9S,9aS,10R)-5 α -[(Acetyloxy)methyl]-5-(benzoyloxy)-octahydro-9-hydroxy-2,2,9-trimethyl-2H-3,9a-methano-1-benzoexepin-7,10-diyl Ester; **2**): Yellow oil. $[\alpha]_D^{20} = +29.0$ ($c = 0.70$, CHCl₃). UV (MeOH): 204, 231, 274. IR (KBr): 3432, 2928, 1727, 1642, 1460, 1238, 1105, 1028, 880, 725. ¹H- and ¹³C-NMR (CDCl₃): Tables 1 and 2. EI-MS: 616 (13.0, M⁺), 493 (8.0, [M – H – BzOH]⁺), 394 (32.0, [M – BzO – MeBuO]⁺), 312 (100), 264 (54), 154 (73). HR-MS: 639.3134 ([M + Na]⁺, C₃₄H₄₈O₁₀Na⁺; calc. 639.3140).

1 β ,6 α ,13-Tris(acetyloxy)-9 β -(cinnamoyloxy)-4 α -hydroxy- β -dihydroagarofuran (= (2E)-3-Phenylprop-2-enoic Acid (3R,5R,5aS,6S,9S,9aS,10R)-6,10-Bis(acetyloxy)-5 α -[(acetyloxy)methyl]-octahydro-9-hydroxy-2,2,9-trimethyl-2H-3,9a-methano-1-benzoexepin-5-yl Ester; **3**): Yellow oil. $[\alpha]_D^{20} = +57.0$ ($c = 5.90$, CHCl₃). UV (MeOH): 218, 223, 281. IR (KBr): 3451, 2933, 1742, 1635, 1370, 1233, 1151, 1032, 880, 712. ¹H- and ¹³C-NMR (CDCl₃): Tables 1 and 2. EI-MS: 410 (8.0, [M – C₆H₅O]⁺), 290 (15.0, [410 – 2AcOH]⁺), 230 (64.0, [290 – AcOH]⁺), 50 (100). FAB-MS: 559 ([M + H]⁺). HR-MS: 559.2534 ([M + H]⁺, C₃₀H₃₉O₁₀⁺; calc. 559.2538).

6 α ,13-Bis(acetyloxy)-9 β -(benzoyloxy)-4 α -hydroxy-1 β -[(2-methylbutanoyl)oxy]- β -dihydroagarofuran (=2-Methylbutanoic Acid (3R,5R,5aS,6S,9S,9aS,10R)-10-(Acetyloxy)-5 α -[(acetyloxy)methyl]-5-(benzoyloxy)-octahydro-9-hydroxy-2,2,9-trimethyl-2H-3,9a-methano-1-benzoexepin-6-yl Ester; **4**): Yellow oil. $[\alpha]_D^{20} = +45.3$ ($c = 3.30$, CHCl₃). UV (MeOH): 203, 231, 274. IR (KBr): 3427, 2928, 2357, 1740, 1632, 1380, 1236, 1145, 1033, 888, 710. ¹H- and ¹³C-NMR (CDCl₃): Tables 1 and 2. EI-MS: 470 (14.0, [M + 2H – MeBuO]⁺), 410 (15.0, [470 – AcOH]⁺), 394 (25.0, [M + 2H – AcO – BzO]⁺), 230 (80), 50 (100). FAB-MS: 575 ([M + H]⁺). HR-MS: 575.2860 ([M + H]⁺, C₃₇H₄₃O₁₀⁺; calc. 575.2851).

Antitumor Assays. Compounds **1–9** were tested for antitumor activity against the following cell lines: P-388, suspension culture of a leukemia neoplasm from a mouse; A-549, monolayer culture of a human-lung carcinoma; HL-60, suspension culture of a leukemia neoplasm from a human; BEL-7402, monolayer culture of a human-liver carcinoma. Cells were maintained in DMEM medium, supplemented with 5% fetal calf serum (FCS), 10-2M sodium hydrogen carbonate and 0.1 g/l penicillin G + 0.1 g/l streptomycin sulfate. The compounds assayed were dissolved in DMSO and tested following the method described by Bergeron *et al.* [19].

H. W. gratefully acknowledges the postdoctoral fellowship awarded by the Zhejiang University, Hangzhou, China, and support by the Lanzhou Medicine Institute, Lanzhou, China.

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Received May 20, 2003